

## Full-length article

# A robust homogeneous binding assay for $\alpha 4\beta 2$ nicotinic acetylcholine receptor<sup>1</sup>

Xin HUI<sup>2</sup>, Jie GAO<sup>2</sup>, Xin XIE<sup>2</sup>, Naoki SUTO<sup>3</sup>, Tsuyoshi OGIKU<sup>3</sup>, Ming-Wei WANG<sup>2,4</sup>

<sup>1</sup>The National Center for Drug Screening, Shanghai Institute of Materia Medica, Shanghai Institutes for Biological Sciences, Chinese Academy of Sciences, Graduate School of Chinese Academy of Sciences, Shanghai 201203, China; <sup>3</sup>Discovery and Pharmacology Research Laboratories, Tanabe Seiyaku Co, 2-50, Kawagishi 2-Chome, Toda, Saitama 335-8505, Japan

## Key words

nicotinic acetylcholine receptor; scintillation proximity assay; high-throughput screening

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<sup>4</sup> Correspondence to Dr Ming-wei WANG.  
Phn 86-21-5080-0598.  
Fax 86-21-5080-0721.  
E-mail mwwang@siniwest.com

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## Abstract

**Aim:** To develop a homogeneous high-throughput screening (HTS) assay based on scintillation proximity assay (SPA) technology for identification of novel  $\alpha 4\beta 2$  nicotinic acetylcholine receptor (nAChR) modulators. **Methods:** Membrane preparation of HEK293 cells expressing  $\alpha 4\beta 2$  nAChR, [<sup>3</sup>H]cytisine and wheat germ agglutinin (WGA)-coupled microbeads were used to develop an HTS assay based on SPA technology. This method was validated against a conventional filter binding approach and applied to large-scale screening of a library containing 32 000 synthetic compounds. Intracellular calcium measurement was carried out to verify the bioactivities of the hits found by the SPA assay. **Results:** IC<sub>50</sub> values of 2 reference compounds (epibatidine and RJR 2403) determined by SPA and filter binding methods were comparable and consistent with those reported elsewhere. A total of 54 compounds, showing more than 60% competitive inhibition on [<sup>3</sup>H]cytisine binding to  $\alpha 4\beta 2$  nAChR, were identified initially following an HTS campaign. Secondary screening confirmed that 17 compounds with novel chemical structures possessed relatively high binding affinity to  $\alpha 4\beta 2$  nAChR ( $K_i < 2 \mu\text{mol/L}$ ). Eight compounds displayed antagonistic effects with >50% inhibition on ABT-594-induced calcium mobilization while none showed any agonist activity. **Conclusions:** This homogeneous binding assay is a highly efficient, amenable to automation and robust tool to screen potential  $\alpha 4\beta 2$  nAChR modulators in an HTS setting. Its application may be expanded to other membrane receptors and ion channels.

## Introduction

Nicotinic acetylcholine receptors (nAChR) are excitatory ligand-gated ion channels mainly distributed in the central and peripheral nervous systems, neuromuscular junctions and adrenal glands<sup>[1]</sup>. The nAChR channel complex is composed of 5 protein subunits, which form a pore that is permeable to Na<sup>+</sup>, K<sup>+</sup>, and Ca<sup>2+</sup>. To date,  $\alpha$ ,  $\beta$ ,  $\gamma$ ,  $\delta$ , and  $\epsilon$  subunits have been cloned, with 9 different  $\alpha$  and 4 varieties of  $\beta$  subunits identified<sup>[2]</sup>. The  $\alpha 4\beta 2$  is the predominant nAChR subtype in the mammalian brain and has a high affinity for nicotine. nAChR consisting of  $\alpha 4$  and  $\beta 2$  subunits modulates neurotransmitter release and plays a direct role in nico-

tine addiction. This group of nAChR was implicated in the pathological pathways of Alzheimer disease, Parkinson disease, schizophrenia, epilepsy and pain<sup>[3]</sup>.

The anti-nociceptive effects of (–)-nicotine have been known for over 60 years. This non-selective nAChR agonist could not be applied as an analgesic agent due to serious adverse events. Although (–)-epibatidine is a highly selective  $\alpha 4\beta 2$  nAChR agonist that displays a more potent analgesic activity than morphine in nociceptive tests<sup>[4]</sup>, toxicities in the cardiovascular and central nervous systems rendered it unsuitable for therapeutic use<sup>[5]</sup>. Therefore, a primary goal in the development of novel analgesic agents based on  $\alpha 4\beta 2$  nAChR is to discover subtype-specific ago-

nists with less liability.

An accurate and robust receptor-binding assay is highly desirable to screen a large collection of chemical entities. Associated techniques are generally based on absorbance, fluorescence and radiometric assays. Scintillation proximity assay (SPA), because of its simplicity and high-throughput nature, has broad applications in measuring receptor-ligand interactions and enzyme reactions. Conventional filter binding assays involve 1 or more separation steps and, thus, are laborious, non-homogeneous and not suitable for automation. SPA technology<sup>[6]</sup>, on the other hand, provides a homogeneous screening approach that does not require post-reaction liquid handling steps and is well suited to automation and high-throughput screening (HTS). Briefly, the receptor is anchored to a scintillant-impregnated microbead. When an isotope (eg, [<sup>3</sup>H]) is brought very close to the microbead by binding to its surface, it activates the scintillant leading to light emission. Because the emitted  $\beta$  particles or auger electrons can only travel short distances in the bulk solution, the microbead preferentially captures electrons from the bound radiolabeled ligand. Therefore, the amount of light emitted from the scintillant in the microbead is directly proportional to the amount of bound radiolabeled ligand. In the present paper, we describe a simple SPA method to assess specific binding properties of various known ligands to  $\alpha 4\beta 2$  nAChR. It was further validated and optimized in the context of HTS for a library containing 32 000 synthetic compounds. More than a dozen compounds with novel chemical structures were found to have relatively high binding affinity to  $\alpha 4\beta 2$  nAChR and 8 of them showed significant antagonist activities in a functional assay.

## Materials and methods

**Reagents** Potassium chloride, sodium phosphate monobasic anhydrous, and magnesium chloride hexahydrate were purchased from Shanghai Chemical Co (Shanghai, China). Aprotinin and leupeptin were purchased from Merck KGaA (Darmstadt, Germany). RJR 2403 and epibatidine were purchased from Sigma-Aldrich (St Louis, MO, USA). [<sup>3</sup>H] Cytisine (37 Ci/mmol) was obtained from Amersham Biosciences UK (Buckinghamshire, UK). FlashBlue™ GPCR beads, Isoplate™ and Filtermat B made of glass fiber were bought from PerkinElmer Life and Analytical Sciences (Boston, MA, USA).

**Cell culture and membrane preparation** The full-length cDNA of the  $\alpha 4$  (GenBank Accession No. U62437) and  $\beta 2$  (GenBank Accession No. U62437) subunits of nAChR were cloned into pcDNA3.1 (Invitrogen, Carlsbad, CA, USA) and

pIRESpuro2 (BD Clontech, Mountain View, CA, USA), respectively. They were subsequently cotransfected into HEK293 cells and clones stably expressing  $\alpha 4\beta 2$  nAChR were selected and characterized by calcium mobilization assay<sup>[7]</sup>. The cells were maintained in Dulbecco's modified Eagle's medium (DMEM) (Sigma) containing 10% fetal calf serum (FCS) in the presence of penicillin (100 mg/L), streptomycin (100 mg/L), G418 (400 mg/L) and puromycin (5 mg/L) at 37 °C in a humidified atmosphere of 5% CO<sub>2</sub>. For membrane preparation, cells were treated with 0.25% trypsin (Sigma) for 5 min and centrifuged at 1000×g for 10 min. The pellets were resuspended in hypotonic buffer [50 mmol/L Tris-HCl, pH 7.5, 5 mmol/L ethylenediaminetetraacetic acid (EDTA)] and homogenized with a BioNeb® cell disruption system (Glas-Col, Terre Haute, IN, USA) followed by centrifugation at 1200×g, 4 °C for 20 min to precipitate nuclear debris. The supernatant was then centrifuged at 20 000×g for 30 min to pellet the membrane. The latter was resuspended in 50 mmol/L Tris-HCl, pH 7.5, 5 mmol/L EDTA with 30% (w/v) sucrose. The protein content was determined using a spectrophotometer (Thermo Electron, Waltham, MA, USA).

**Filter binding assay** Various amounts of the above membrane receptor preparation, 3 nmol/L [<sup>3</sup>H]cytisine and different concentrations of known ligands to  $\alpha 4\beta 2$  nAChR were added to the binding buffer (50 mmol/L Tris HCl, 120 mmol/L NaCl, 5 mmol/L KCl, 1 mmol/L MgCl<sub>2</sub>, 2.5 mmol/L CaCl<sub>2</sub>, pH 7.4) to give a final volume of 0.1 mL. After incubation at 4 °C for 8 h, the membrane fraction was harvested on the Filtermat B presoaked with a solution consisting of the washing buffer (50 mmol/L Tris HCl, pH 7.4) and 0.5% (v/v) polyethyleneimine. The Filtermat B was washed 3 times with the washing buffer and dried at 55 °C for 45 min before counting on a Microbeta scintillation counter (PerkinElmer). Non-specific binding activity was determined in the presence of 2  $\mu$ mol/L epibatidine.

**SPA binding assay** Various amounts of the above membrane receptor preparation, 3 nmol/L [<sup>3</sup>H]cytisine, FlashBlue™ GPCR beads (62.5  $\mu$ g/well) and different concentrations of known ligands to  $\alpha 4\beta 2$  nAChR were added to the binding buffer to give a final volume of 0.1 mL. The plates were incubated at 4 °C for 12 h and centrifuged for 3 min at 2500×g before counting on the Microbeta scintillation counter.

**Calcium mobilization assay** The above cells expressing  $\alpha 4\beta 2$  nAChR were detached and loaded with 5  $\mu$ mol/L Fluo-4 (Molecular Probes, Eugene, OR, USA) in culture medium supplemented with 2.5 mmol/L probenecid (Sigma) for 45 min. They were then washed twice with solB buffer containing: 10 mmol/L HEPES, 5 mmol/L NaCl, 5 mmol/L KCl, 2 mmol/L

CaCl<sub>2</sub>, 140 mmol/L *N*-methyl-D-glucamine and 10 mmol/L glucose (pH 7.4). The cells were resuspended in solB buffer, plated onto 96-well plates at a density of 60 000 cells in 80 μL per well, and reattached by centrifugation. Effects of the hit compounds on calcium influx were analyzed by FlexStation™ (Molecular Devices, Sunnyvale, CA, USA) with excitation wavelength 485 nm and emission wavelength 525 nm. An α4β2 nAChR agonist, ABT-594<sup>[8]</sup>, was used as a positive control.

**HTS campaign** The compound library used for screening consisted of 32 000 pure synthetic compounds. A 10-compound pool per well mix was applied to the primary screening, with an average concentration of 7 μmol/L for each compound dissolved in 100% dimethylsulphoxide (Me<sub>2</sub>SO) solution. This matrix system maximizes the advantage of HTS and allows duplicate screening of each compound<sup>[9]</sup>. In each 96-well Isoplate™, 16 wells were used as positive controls (epibatidine) and samples showing greater than 60% inhibition were considered as “hits”.

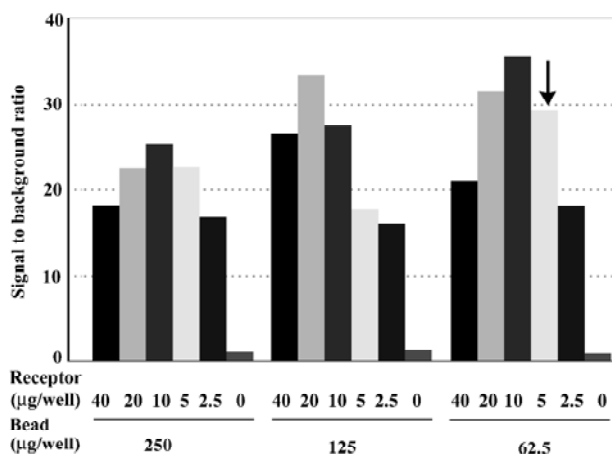
**Data analysis** Data were analyzed using GraphPad Prism software (GraphPad, San Diego, CA, USA). Non-linear regression analyses were performed to generate dose-response curves. *K<sub>i</sub>* values were calculated from IC<sub>50</sub> using the equation of Cheng and Prusoff<sup>[10]</sup>.

$$K_i = \frac{IC_{50}}{1 + \frac{[radioligand]}{K_d}}$$

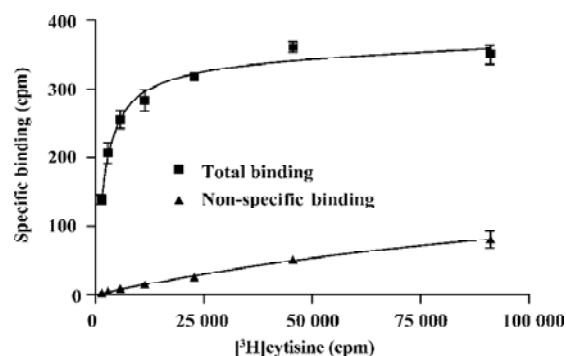
## Results

**Assay optimization** For HTS purposes, 96-well microtiter plates were used and reaction volume was adjusted to 100 μL/well. At a concentration equivalent to the previously reported *K<sub>d</sub>* value (~0.5 nmol/L)<sup>[11]</sup>, more than 25% of [<sup>3</sup>H]cytisine added was bound to the receptor (data not shown). To avoid excessive ligand depletion, the concentration was increased to 3 nmol/L, in which the bound form accounted for less than 10%. The best signal-to-background (S/B) ratio was observed in a matrix experiment when 62.5 μg/well microbeads and 10 μg/well membrane preparation were used. However, 5 μg/well was selected for the HTS campaign to conserve the membrane preparation. Under these assay conditions, a sound S/B ratio (~30) was achieved (Figure 1). Me<sub>2</sub>SO, at concentrations below 3%, did not affect the assay performance (data not shown).

**Binding saturation** A serial titration of [<sup>3</sup>H]cytisine was prepared to study binding characteristics of the SPA assay (Figure 2). The *B<sub>max</sub>* and *K<sub>d</sub>* values calculated from the SPA and filter binding assays were strikingly similar: 2.26 pmol/mg vs 2.31 pmol/mg and 0.65 nmol/L vs 0.70 nmol/L,



**Figure 1.** Assay optimization. Different concentrations of FlashBlue™ GPCR beads and membrane preparation were mixed in the presence of 3 nmol/L of [<sup>3</sup>H]cytisine. Unlabeled epibatidine was included at a concentration of 2 μmol/L to determine non-specific binding. The signal was measured after 12 h incubation using a MicroBeta scintillation counter. The arrow indicates the conditions to be optimised.

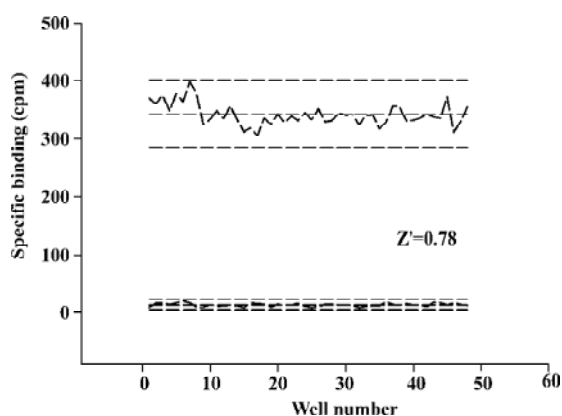


**Figure 2.** Saturation study in the scintillation proximity assay. A series titration of [<sup>3</sup>H]cytisine was made to determine receptor binding characteristics in the presence and absence of 2 μmol epibatidine. *n*=3. Mean±SD.

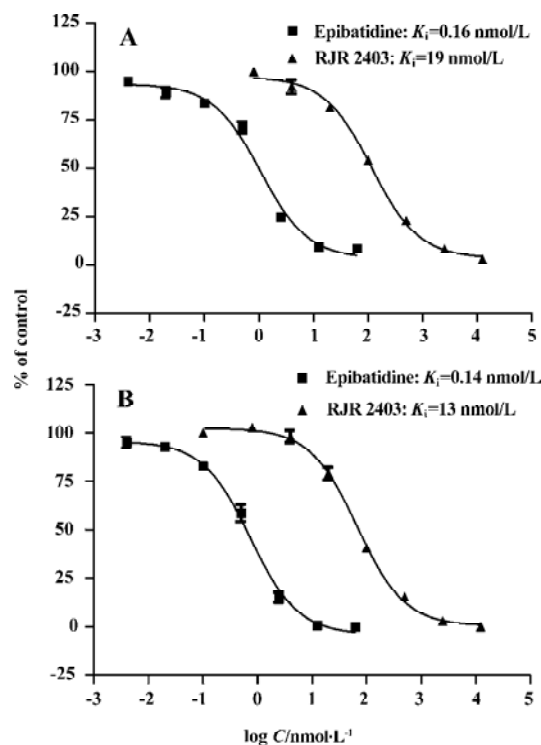
respectively.

**Assay performance** As shown in Figure 3, the average *Z'* value for the SPA binding assay was 0.78 with a S/B ratio of 29, indicating that the system was adequately optimized for HTS. Assay stability was evaluated by incubating the plates overnight at 4 °C and both the *Z'* factor and the S/B ratio remained unchanged. Two known α4β2 nAChR ligands, namely, epibatidine and RJR2403, were used to compare the 2 assay methods and the binding affinities measured were within the same range (Figure 4).

**HTS campaign** Of the 32 000 samples initially screened, 54 hits (0.17%) showing greater than 60% competitive inhibition on [<sup>3</sup>H]cytisine binding to α4β2 nAChR were discov-

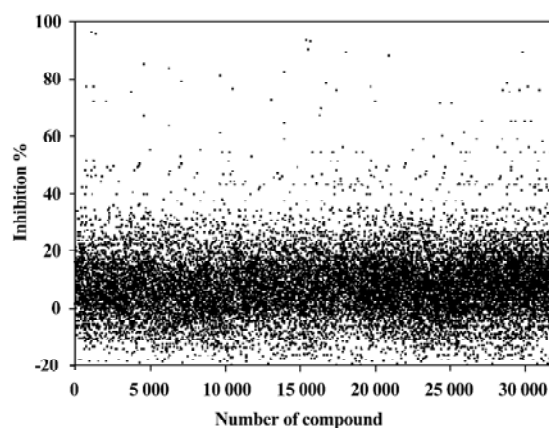


**Figure 3.** Z' factor determination. Assays were performed at optimized conditions (62.5  $\mu$ g beads+5  $\mu$ g membrane preparation). Forty-eight replicates of total and non-specific signals were studied. Dashed lines indicate means and mean $\pm$ 3SD of 48 data points.



**Figure 4.** Dose-response curves of epibatidine and RJR 2403 measured by the (A) scintillation proximity assay or (B) filter binding method from which respective  $K_d$  values were calculated.  $n=3$ . Mean $\pm$ SD.

ered (Figure 5). Secondary (single compound per well) screening confirmed that 17 of the above hits displayed consistent inhibitory effects with  $K_i$  values below 2  $\mu$ mol/L. These active compounds are of different chemical structures including thiophene, piperidine, azole and other types of heterocyclic derivatives. The HTS campaign was high quality in na-



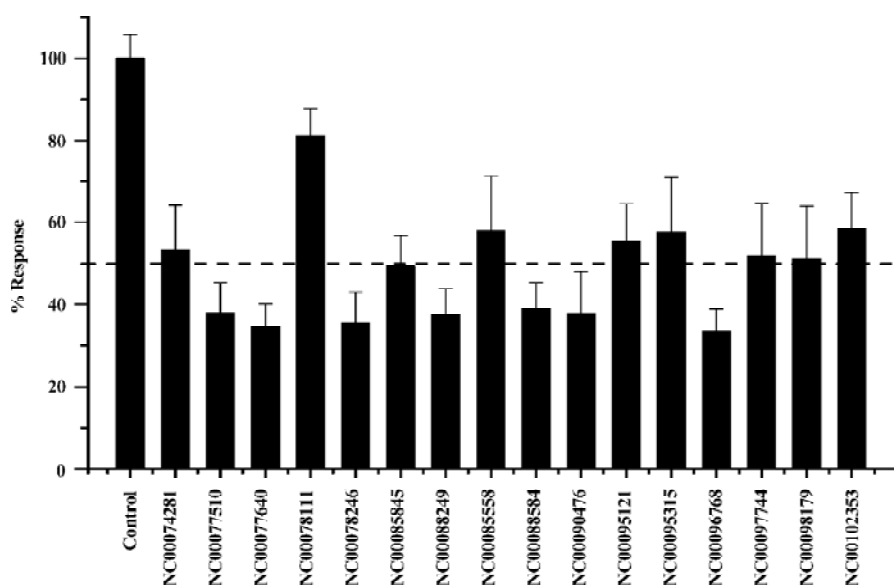
**Figure 5.** High-throughput screening of 32 000 compounds using the scintillation proximity assay. Results are expressed as percentage inhibition of [ $^3$ H]cytisine binding to  $\alpha$ 4 $\beta$ 2 nicotinic acetylcholine receptor.

ture with a Z' factor and S/B ratio equaling 0.73 and 29, respectively. The bioactivities of the confirmed hits were further evaluated with a functional assay. Eight compounds displayed antagonistic effects with >50% inhibition at 6.7 mg/L on ABT-594-induced calcium mobilization while none showed any agonist activity (Figure 6).

## Discussion

Taking advantage of our prior experience in the development of a SPA-based nuclear receptor binding HTS method<sup>[12]</sup>, efforts were made to expand the knowledge to membrane receptors such as  $\alpha$ 4 $\beta$ 2 nAChR described in this paper. Although similar SPA approaches have been employed to study ligand-binding characteristics of a variety of G protein-coupled receptors<sup>[6,13-15]</sup>, its application in  $\alpha$ 4 $\beta$ 2 nAChR is novel and may be expandable to other ligand-gated ion channels.

When developing a SPA-based receptor-binding assay, many factors have to be considered in order to maximize the S/B ratio. The optimization procedure may include scintillant-impregnated microbead selection, assay buffer and volume determination, or protein/bead ratio verification. While the assay buffer could be readily transferred from a conventional filter binding assay, the amount of receptor protein and its relative ratio to the quantity of beads used are critical to assay performance. An adequate amount of beads is required to saturate the protein in order to achieve a maximal binding signal. However, excessive beads will lead to undesired exposure of bead surface to non-specific ligand binding. Therefore, the amount of beads applied should be kept to a minimum where protein saturation is still achievable.



**Figure 6.** Inhibition of  $\alpha 4\beta 2$  nicotinic acetylcholine receptor agonist ABT-594-induced calcium mobilization by 16 hit compounds (6.7 mg/L). One compound was not included due to auto-fluorescence. Cells were pretreated with test compounds or dimethylsulphoxide (control) for 2 min and calcium influx was stimulated by ABT-594 (166 nmol/L).  $n=3$ . Mean $\pm$ SEM.

On the other hand, stable signal is highly dependent on protein stability and its coating ability on the beads. In the assay system reported here, no signal shift was observed for more than 48 h, indicating a very stable interaction among various reagents.

It was learned from other membrane receptor screening settings that assay sensitivity can be improved by choosing a radiolabeled ligand concentration at or below its  $K_d$  to permit effective competition by an unlabeled ligand<sup>[14]</sup>. In the present study, we selected a relatively higher concentration of [<sup>3</sup>H]cytisine (ie, 3 nmol/L) as opposed to the  $K_d$  value (0.6 nmol/L) of unlabeled ligand. This arrangement allowed us to accurately measure  $K_i$  values of the 2 reference compounds as well as to confirm hits identified from the HTS campaign, while keeping [<sup>3</sup>H]cytisine bound to the receptor to a minimum (<10%). Conceivably, such a practice would compromise assay sensitivity thereby reducing the hit rate. Considering our initial intention of finding hits with  $K_i$  values below 2  $\mu$ mol/L from primary screening, this purpose was well served as evidenced by a confirmed hit rate of 0.05% and the identification of 1 highly active compound (23 nmol/L; data not shown). In addition, the bioactivities of the hits discovered by the SPA method were confirmed with a cell-based functional assay where 8 compounds were found to demonstrate significant antagonistic effects on ABT-594-induced calcium mobilization.

In comparison with conventional filter binding techniques, the SPA method omits steps such as pre-coating, pre-incubation, separation and excessive washing, and thus, simplifies the assay protocol, mitigating labor intensity and

reducing systemic error. It is also readily adaptable to HTS and automation, as demonstrated in this study. When deduced to practice, both approaches yielded similar  $K_i$  values for 2 known  $\alpha 4\beta 2$  nAChR ligands (epibatidine and RJR2403), which are not only consistent with those reported elsewhere<sup>[16]</sup> but also exhibited the same affinity rank order and  $pK_i$  features (in agreement within 1/2 log unit). In addition, the cost and amount of wastage are significantly less if SPA technology is employed. This advantage could be further explored by utilizing high-density plate formats (eg, 384-well plate) where filtering assays clearly show their limitations.

The  $Z'$  factor is a useful tool for evaluating bioassay qualities<sup>[17]</sup>. In general, a  $Z'$  value above 0.5 suggests that an assay is robust enough for HTS. The SPA system described herein consistently displayed a  $Z'$  value between 0.73 and 0.78. This, and in conjunction with other parameters such as  $S/B$  ratio,  $B_{max}$  and  $K_d$  values, indicate that the assay is high quality in nature.

In summary, a simple and SPA-based HTS binding assay was developed and validated for identification of compounds with specificity and functionality for  $\alpha 4\beta 2$  nAChR. Receptor coating onto the beads and ligand binding are achieved in 1 mixing step and the procedure is homogeneous. Its application may be expanded to other membrane receptors and ion channels.

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